

Catalytic Asymmetric Construction of Vicinal Tetrasubstituted Stereocenters by the Mannich Reaction of Linear α -Substituted Monothiomalonates with Isatin *N*-Boc Ketimines

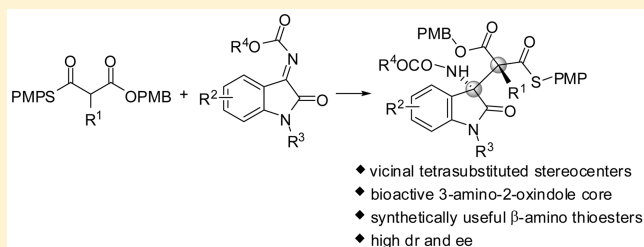
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S Supporting Information

ABSTRACT: A highly diastereo- and enantioselective method for the construction of vicinal tetrasubstituted stereocenters through the first catalytic asymmetric Mannich reaction of linear α -substituted MTMs with isatin *N*-Boc ketimines has been developed. This protocol provides atom-economic synthesis of less accessible 3-aminooxindoles bearing vicinal tetrasubstituted stereocenters. Notably, it also constitutes the first example of stereoselective synthesis of β -amino thioesters bearing vicinal tetrasubstituted stereocenters.



INTRODUCTION

Stereoselective construction of vicinal tetrasubstituted stereocenters is of great importance,¹ because such structural motifs are present in numerous natural products and bioactive compounds.² However, although substrate-directed diastereoselective methods for the generation of vicinal tetrasubstituted stereocenters have been well documented, catalytic asymmetric approaches to construct these highly congested stereochemical dyads remain a formidable challenge due to low reactivity caused by steric repulsion and the introduction of requisite diastereocontrol. To date, only a few catalytic asymmetric approaches have been reported to generate contiguous tetrasubstituted stereocenters via a single synthetic operation. Among these approaches, cycloadditions³ and alkylation reactions⁴ are most commonly used. In recent years, the catalytic asymmetric nucleophilic 1,2-addition of trisubstituted carbon nucleophiles to ketones and ketimines⁵ has emerged as a potential method for constructing vicinal tetrasubstituted stereocenters because of its atom economy⁶ and incorporation of the alcohol or amine functionality. However, the reported examples generally employ cyclic nucleophiles or both vicinal tetrasubstituted stereocenters are formed at a constrained cycle. To address these limitations, we initiated studies investigating the catalytic asymmetric Mannich reaction of ketimines employing linear α -substituted monothiomalonates to construct challenging vicinal tetrasubstituted stereocenters.

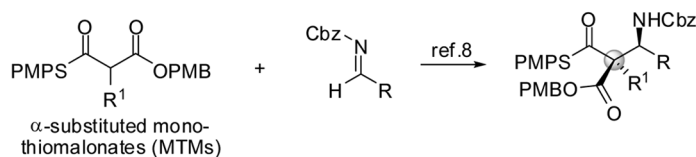
A new class of linear trisubstituted carbon nucleophiles, α -substituted monothiomalonates (MTMs), have recently introduced by Wennemers.^{7a} The synthetic potential of α -substituted MTMs has been demonstrated in the asymmetric Michael addition of nitroolefins.⁷ Very recently, Wennemers and co-workers disclosed the first asymmetric 1,2-addition of α -

substituted MTMs to aldimines (Scheme 1a).⁸ This reaction provides an elegant method for the enantioselective synthesis of β -amino thioesters, which are important building blocks for the synthesis of bioactive molecules. However, they noted that although α -substituted MTMs reacted smoothly with *N*-Cbz-protected aldimines, this class of nucleophiles did not react with the corresponding *N*-Boc-protected aldimines because of steric restraints.⁸ Thus, it is not surprising that there are no reports challenging the difficulty involved in the enantioselective addition of α -substituted MTMs to ketimines, even though such a reaction, if successful, would provide an attractive atom-economic method for vicinal tetrasubstituted stereocenters. Compared to aldimines, ketimines are typically less reactive due to steric restraints. Additionally, this process is complicated further because a second vicinal tetrasubstituted center is present in the synthetic target. However, the importance of enantioenriched β -amino thioesters bearing contiguous tetrasubstituted stereocenters combined with the virtual absence of reports for their stereoselective synthesis encouraged our exploration of this unprecedented Mannich reaction⁹ of linear α -substituted MTMs with *N*-Boc ketimines. Here, isatin *N*-Boc ketimines were chosen as the substrates because the resulting β -amino thioesters have a 3-aminooxindole backbone, which is the core structure in numerous natural products and bioactive molecules.^{10,11}

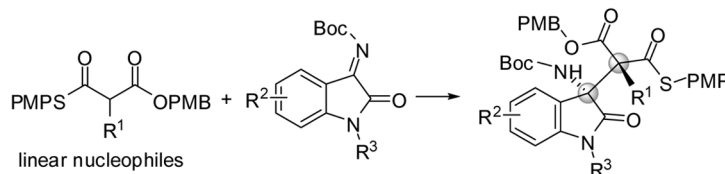
The catalytic asymmetric nucleophilic 1,2-addition to isatin-derived ketimines has received increasing attention^{12,13} because of the importance of 3-aminooxindoles. However, despite remarkable advances, most reports generate 3-aminooxindole

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Scheme 1. Construction of Tetrasubstituted Stereogenic Centers in the Addition of α -Substituted MTMs to Imines(a): Addition of α -substituted MTMs to *N*-Cbz aldimines (previous work)

♦ α -Substituted MTMs did not react with *N*-Boc aldimines due to steric restraints.

(b) Addition of α -substituted MTMs to *N*-Boc ketimines (this work)

- ♦ vicinal tetrasubstituted stereocenters
- ♦ bioactive 3-amino-2-oxindole core
- ♦ synthetically useful β -amino thioesters
- ♦ high dr and ee

products containing a single stereocenter¹² with only a few examples furnishing products containing vicinal quaternary and tertiary stereocenters with good stereoselectivities.¹³ Catalytic asymmetric nucleophilic 1,2-additions to isatin-derived ketimines targeting 3-aminoxindoles bearing vicinal tetrasubstituted stereocenters are rare. During the preparation of our manuscript, Feng et al. reported a metal-catalyzed asymmetric addition of linear silyl ketene imine to isatin-derived ketimine.¹⁴ However, despite its elegance, to achieve high diastereo- and enantioselectivity, lower temperature ($-45\text{ }^{\circ}\text{C}$) was required. To date, the organocatalytic asymmetric nucleophilic 1,2-additions of linear nucleophiles to isatin-derived ketimines targeting 3-aminoxindoles bearing vicinal tetrasubstituted stereocenters remains to be developed.

RESULTS AND DISCUSSION

We began our study by evaluating a variety of bifunctional tertiary amine–thiourea organocatalysts¹⁵ (Figure 1) for the

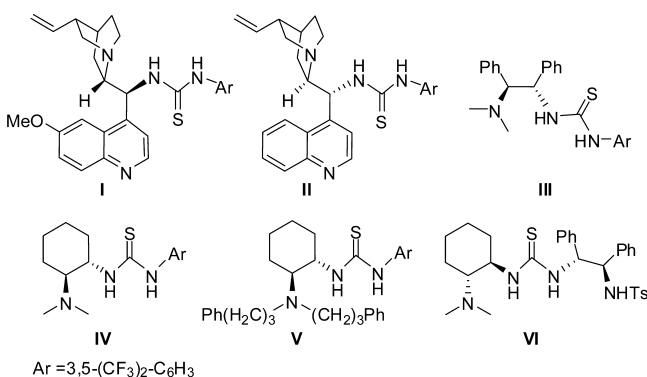
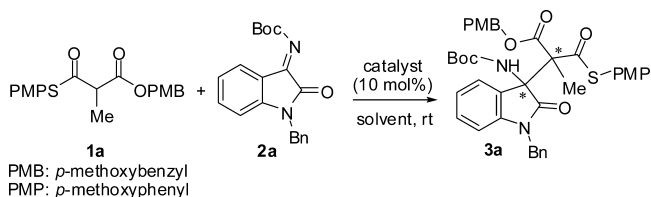


Figure 1. Structures of catalysts screened in this study.

model reaction of **1a** and **2a** at room temperature, and the results are summarized in Table 1. It was found that the catalyst structures have remarkable effects on the stereochemical results. Using quinine-derived thiourea **I**, product **3a** was obtained with high enantioselectivity (98% ee) but moderate diastereoselectivity (5:1 dr) (entry 1). However, the use of cinchonine-derived thiourea **II** provided product **3a** with poor diastereo-

Table 1. Optimization of the Reaction Conditions^a

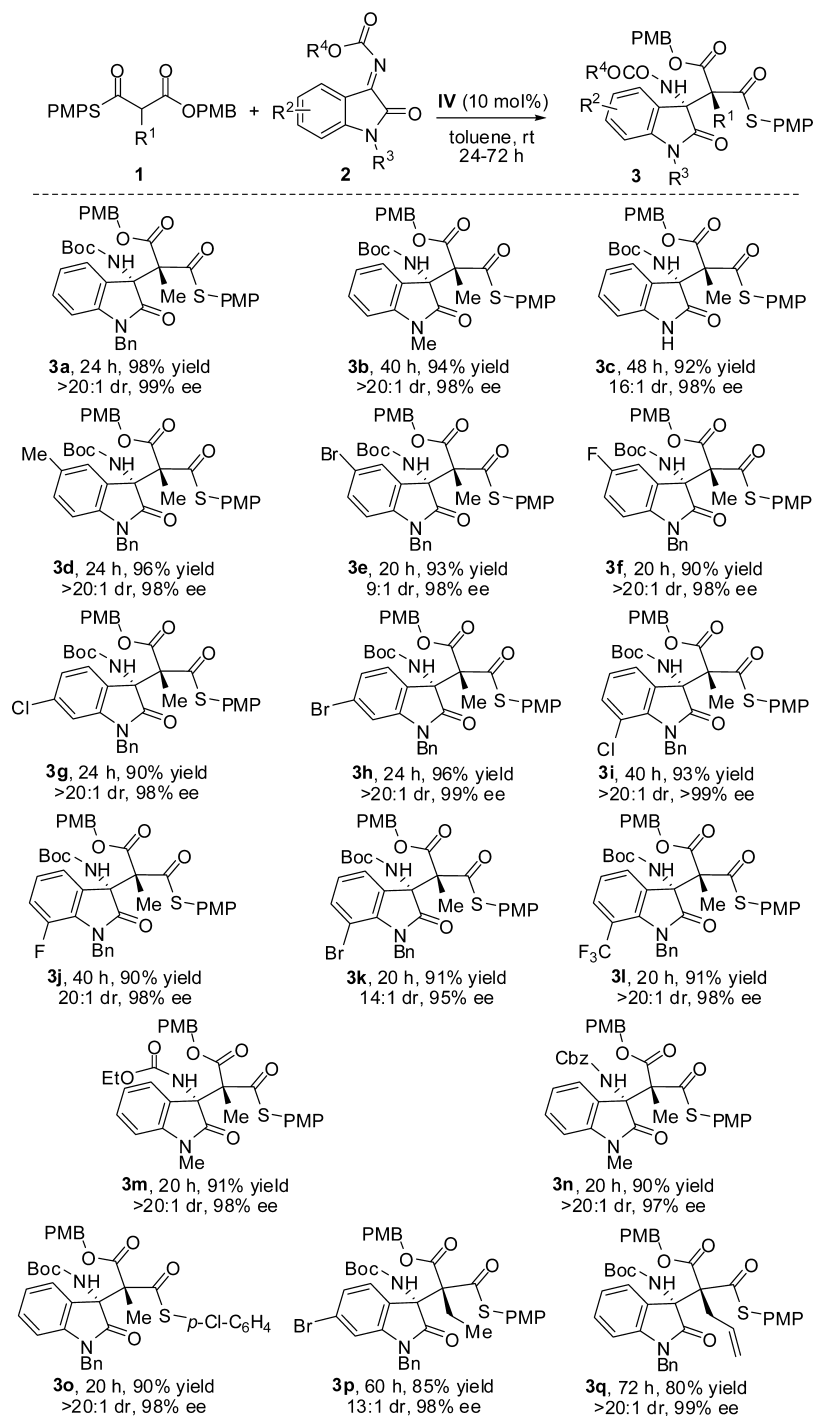


entry	catalyst	solvent	time (h)	yield (%) ^b	dr ^c	ee (%) ^c
1	I	toluene	40	75	5:1	98
2	II	toluene	48	70	1:1	-25
3	III	toluene	48	82	4:1	-19
4	IV	toluene	20	98	>20:1	-99
5	V	toluene	48	68	2:1	-7
6	VI	toluene	66	85	4:1	92
7	IV	CHCl ₃	70	92	6:1	96
8	IV	MeCN	72	75	4:1	92
9	IV	EtOAc	60	95	7:1	95

^aReactions was performed with **1a** (0.11 mmol) and **2a** (0.1 mmol) in the presence of catalyst (10 mol %) at room temperature. ^bIsolated yield. ^cDetermined by chiral HPLC.

and enantioselectivity (1:1 dr and 25% ee) (entry 2). Diphenylethane-1,2-diamine derived tertiary amine thiourea **III** gave poor enantioselectivity (19% ee) and moderate diastereoselectivity (4:1 dr) (entry 3). When cyclohexane-1,2-diamine-derived tertiary amine thiourea **IV** was used, the desired product was obtained in 98% yield with 99% ee and >20:1 dr (entry 4). This is useful as both enantiomers of the product could be obtained with excellent enantioselectivity values. To gain further insights into the effects of the catalyst structure on the reaction, we examined cyclohexane-1,2-diamine-derived tertiary amine thiourea catalysts **V** and **VI**. Interestingly, bulky catalyst **V** gave poor diastereo- and enantioselectivity (entry 5) whereas catalyst **VI** bearing multiple hydrogen bond donors afforded good enantioselectivity but moderate diastereoselectivity (entry 6). Then, solvent effects were examined. Whereas the solvents had little influence on enantioselectivity, they had remarkable effects on diastereoselectivity (entries 4 and 7–9). Ultimately, toluene proved to be the optimal solvent (entry 4).

Scheme 2. Substrate Scope



Upon the optimized conditions being determined, the substrate scope was investigated. The Mannich reaction with a range of isatin-derived *N*-Boc ketimines proceeded smoothly to provide the products in excellent yields and diastereo- and enantioselectivities (Scheme 2, 3a–l). The ketimines, with protecting groups such as Cbz and $\text{CH}_3\text{CH}_2\text{OCO}$ -, also worked very well (3m and 3n).¹⁶ Notably, the ketimine without the substituted group at the 1-position could also react well with α -methyl-substituted MTM to afford Mannich product 3c in 92% yield with 16:1 dr and 98% ee. Other MTMs bearing ethyl and allyl substituents at the α -position were also tolerated well, and the products 3p and 3q were obtained in high yields with

diastereoselectivities of 13:1 to >20:1 and enantioselectivities of 98 to >99% ee. MTM with a *p*-chlorophenyl thioester also worked well to provide product 3o in 90% yield with >20:1 dr and 98% ee.

The absolute and relative configurations of all of the Mannich products (3) were unambiguously assigned on the basis of X-ray crystallographic analysis of 3h (Figure 2).¹⁷

On the basis of the X-ray structure of 3h and the work by Wennemers,⁸ a proposed stereochemical model is illustrated in Figure 3. It involves coordination of the MTM to the urea moiety of the catalyst¹⁸ and addition of the MTM enolate from the *Re* face to the *Re* face of the imine (Figure 3, left). An

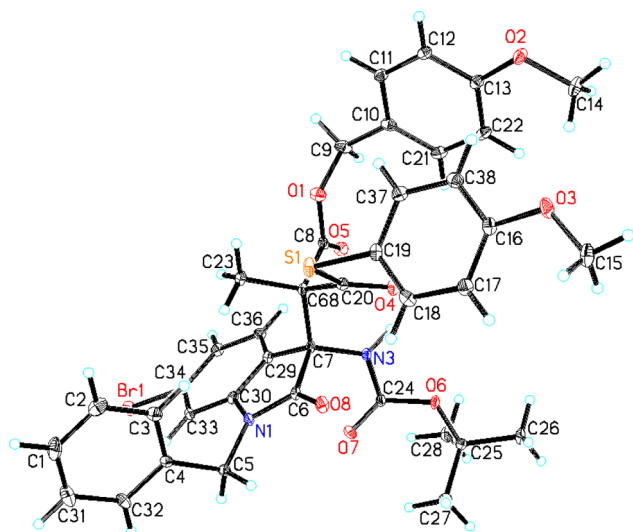


Figure 2. X-ray crystal structure of **3h** (displacement ellipsoids are drawn at the 30% probability level).

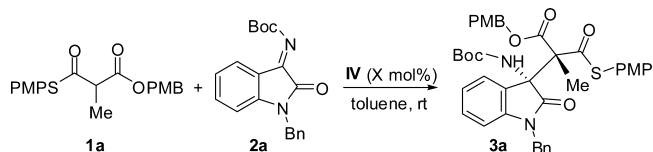
alternative *Si-Si* face approach is less likely due to unfavorable steric interactions (Figure 3, right).

Encouraged by the unexpected high efficiency of this asymmetric Mannich process, we further examined reactions with low catalyst loading, and the results are summarized in Table 2. When the catalyst loading was decreased from 10 to 2 mol %, remarkably high yield (94%) and excellent diastereo- and enantioselectivity (>20:1 dr and >99% ee) were still obtained, although a long reaction time was necessary (entries 1–3). However, a further decrease in the catalyst loading to 1 mol % led to a drop in diastereoselectivity (10:1 dr) albeit in excellent enantioselectivity (>99% ee) and high yield (92%) (entry 4).

CONCLUSION

In summary, we have developed a highly diastereo- and enantioselective method for the construction of vicinal tetrasubstituted stereocenters through the first Mannich reaction of linear α -substituted MTMs with isatin *N*-Boc ketimines. The catalyst structure had a remarkable influence on the stereoselectivity. By using cyclohexane-1,2-diamine-derived tertiary amine-thiourea catalyst **IV**, less accessible chiral 3-aminooxindoles bearing vicinal tetrasubstituted stereocenters were obtained in an atom-economic manner in high yields (up

Table 2. Reaction with Low Catalyst Loading^a



entry	X (mol %)	time (h)	yield (%) ^b	dr ^c	ee (%) ^c
1	10	20	98	>20:1	99
2	5	47	97	>20:1	>99
3	2	96	94	>20:1	>99
4	1	96	92	10:1	>99

^aThe reactions was performed with **1a** (0.11 mmol) and **2a** (0.1 mmol) in the presence of **IV** (X mol %) at room temperature. ^bIsolated yield. ^cDetermined by chiral HPLC analysis.

to 98% yield) and remarkably high stereoselectivities (up to >20:1 dr and 99% ee). The present reaction also constitutes the first example of stereoselective synthesis of β -amino thioesters bearing vicinal tetrasubstituted stereocenters.

EXPERIMENTAL SECTION

¹H and ¹³C NMR spectra were recorded at 300 and 400 MHz, respectively, on a spectrophotometer. Chemical shifts (δ) are expressed in ppm, and *J* values are given in Hz. The enantiomeric excess was determined by chiral HPLC with *n*-hexane and *i*-propanol as eluents. High resolution mass spectrometry (HRMS) was recorded on a spectrometer using a time-of-flight (TOF) analyzer. Optical rotations were measured on a polarimeter. All chemicals and solvents were used as received without further purification unless otherwise stated. Flash column chromatography was performed on silica gel (200–300 mesh).

General Procedure for the Catalytic Asymmetric Mannich Reactions of α -Substituted MTMs **1 with Isatin *N*-Boc Ketimines **2**.** To a solution of *N*-Boc ketimines **2** (0.1 mmol) and catalyst **IV** (0.01 mmol, 0.1 equiv) in toluene (0.25 mL) was added α -substituted MTMs **1** (0.11 mmol, 1.1 equiv). The reaction mixture was stirred at room temperature until the reaction was complete (monitored by TLC). The solvent was removed under reduced pressure, and the crude product was purified by flash silica gel chromatography (ethyl acetate/petroleum ether = 1:5) to afford product **3**.

(R)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)-amino)-2-oxindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3a**). White solid (70.4 mg, 98% yield). Mp 154–156 °C. [α]_D²⁰ –9.6 (c 1.0 CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.36 (d, *J* = 7.2 Hz, 2H), 7.25–7.21 (m, 2H), 7.18–7.13 (m, 5H), 7.07 (t, *J* = 8 Hz, 1H), 7.01–6.99 (m, 2H), 6.85–6.78 (m, 5H), 6.58 (d, *J* = 7.6 Hz, 1H), 5.05 (m, 2H), 4.92–4.88 (m, 1H), 4.77–4.74 (m, 1H), 3.75

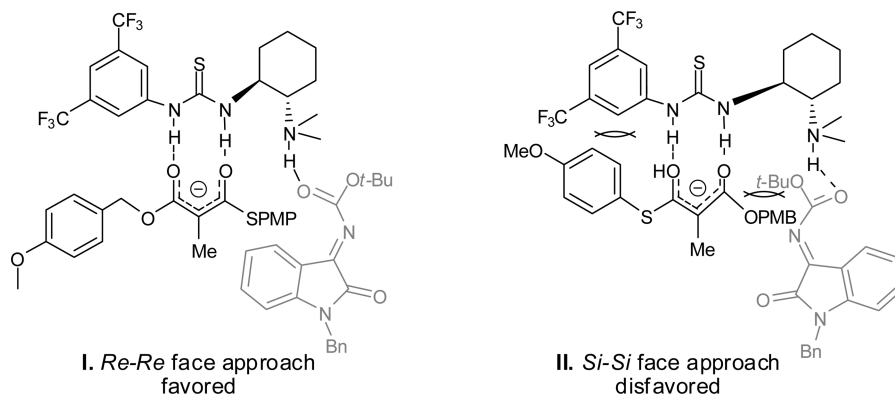


Figure 3. Proposed stereochemical model.

(s, 3H), 3.73 (s, 3H), 1.49 (s, 3H), 1.17 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.3, 173.8, 168.1, 161.0, 159.8, 153.8, 144.2, 136.5, 135.9, 130.3, 129.3, 128.7, 127.8, 127.5, 126.9, 124.2, 122.6, 117.6, 115.0, 113.9, 108.9, 79.9, 68.0, 65.6, 63.1, 55.4, 55.3, 44.6, 28.2, 15.8. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{40}\text{N}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 719.2397; found, 719.2398. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 10:90, flow rate = 0.5 mL/min, λ = 254 nm): t_{minor} = 30.1 min, t_{major} = 44.0 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-3-((*tert*-Butoxycarbonyl)amino)-1-methyl-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3b**). White solid (60.4 mg, 94% yield). Mp 159–161 °C. $[\alpha]_{\text{D}}^{20}$ +10.6 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.22–7.19 (m, 3H), 7.14 (d, J = 8 Hz, 2H), 6.98 (d, J = 7.2 Hz, 1H), 6.94 (s, 1H), 6.87–6.80 (m, 5H), 6.67 (d, J = 8 Hz, 1H), 5.12–5.05 (m, 2H), 3.74 (s, 6H), 3.11 (s, 3H), 1.49 (s, 3H), 1.13 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.2, 173.6, 168.0, 160.9, 159.9, 153.7, 144.8, 136.5, 130.4, 129.5, 127.6, 126.9, 124.1, 122.6, 117.6, 115.0, 113.9, 107.9, 79.8, 68.1, 65.6, 63.0, 55.4, 55.3, 28.1, 26.5, 15.7. HRMS (ESI-TOF): calcd for $\text{C}_{33}\text{H}_{36}\text{N}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 643.2084; found, 643.2082. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 15:85, flow rate = 0.4 mL/min, λ = 254 nm): t_{major} = 27.0 min, t_{minor} = 34.3 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3c**). White solid (57.8 mg, 92% yield). Mp 132–133 °C. $[\alpha]_{\text{D}}^{20}$ –17.1 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.76 (s, 1H), 7.22–7.20 (m, 3H), 7.14 (d, J = 8 Hz, 1H), 7.09 (t, J = 7.6 Hz, 1H), 6.97–6.95 (m, 2H), 6.85–6.80 (m, 5H), 6.63 (d, J = 7.6 Hz), 5.14–5.07 (m, 2H), 3.74 (s, 6H), 1.56 (s, 3H), 1.18 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.4, 174.8, 168.0, 161.0, 159.9, 153.9, 142.0, 136.5, 130.4, 129.4, 128.0, 126.9, 124.5, 122.5, 117.5, 115.0, 113.9, 109.9, 80.2, 68.1, 65.9, 62.9, 55.4, 55.3, 28.2, 15.6. HRMS (ESI-TOF): calcd for $\text{C}_{32}\text{H}_{34}\text{N}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 629.1928; found, 629.1930. HPLC (Chiralpak AS-H, *i*-propanol/*n*-hexane = 15:85, flow rate = 0.6 mL/min, λ = 254 nm): t_{minor} = 23.2 min, t_{major} = 51.2 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-5-methyl-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3d**). Light yellow solid (70.3 mg, 96% yield). Mp 161–163 °C. $[\alpha]_{\text{D}}^{20}$ –8.2 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.43 (d, J = 8 Hz, 2H), 7.32–7.24 (m, 7H), 7.12 (s, 1H), 6.93 (d, J = 8 Hz, 3H), 6.87 (d, J = 8.4 Hz, 2H), 6.82 (s, 1H), 6.53 (d, J = 8 Hz, 1H), 5.17–5.10 (m, 2H), 4.96–4.92 (m, 1H), 4.85–4.82 (m, 1H), 3.83 (s, 3H), 3.80 (s, 3H), 2.17 (s, 3H), 1.53 (s, 3H), 1.26 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.5, 173.6, 167.9, 161.0, 159.8, 153.7, 141.7, 136.5, 136.0, 132.0, 130.3, 129.6, 128.6, 127.8, 127.5, 126.9, 124.9, 117.6, 115.0, 113.9, 108.7, 79.8, 68.0, 65.8, 63.0, 55.4, 55.3, 44.5, 28.2, 21.2, 15.7. HRMS (ESI-TOF): calcd for $\text{C}_{40}\text{H}_{42}\text{N}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 733.2554; found, 733.2558. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 5:95, flow rate = 0.5 mL/min, λ = 254 nm): t_{minor} = 51.6 min, t_{major} = 95.0 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-5-bromo-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3e**). White solid (74.0 mg, 93% yield). Mp 171–172 °C. $[\alpha]_{\text{D}}^{20}$ –9.8 (c 2.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.44–7.42 (m, 2H), 7.36–7.32 (m, 2H), 7.30–7.22 (m, 7H), 7.01 (s, 1H), 6.97 (d, J = 8.4 Hz, 2H), 6.90 (d, J = 8.4 Hz, 2H), 6.52 (d, J = 8.0 Hz, 1H), 5.11 (m, 2H), 4.97–4.83 (m, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 1.63 (s, 3H), 1.30 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 197.0, 173.2, 167.8, 161.1, 159.9, 153.7, 143.2, 136.6, 135.4, 132.1, 130.5, 130.3, 129.8, 128.7, 127.8, 127.7, 127.4, 126.6, 117.2, 115.3, 115.1, 114.0, 110.4, 80.2, 68.2, 65.2, 63.1, 55.4, 55.3, 44.6, 28.2, 15.9. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{BrN}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 797.1502; found, 797.1505. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 15:85, flow rate = 0.4 mL/min, λ = 254 nm): t_{minor} = 34.0 min, t_{major} = 35.6 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-5-fluoro-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3f**). White solid (66.2 mg, 90% yield). Mp 164–165 °C. $[\alpha]_{\text{D}}^{20}$ +21.0 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.45–7.41 (m, 2H), 7.36–7.32 (m, 2H), 7.30–7.25 (m, 5H), 7.05 (s, 1H), 6.96 (d, J = 7.6 Hz, 2H), 6.93–6.83 (m, 4H), 6.58–6.55 (m, 1H), 5.13 (m, 2H), 4.99–4.87 (m, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 1.59 (s, 3H), 1.30 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ

196.7, 173.6, 167.8, 161.0, 159.9, 153.7, 140.1, 136.5, 135.6, 130.4, 128.7, 127.8, 127.6, 126.7, 117.3, 115.6, 115.4, 115.0, 114.0, 112.8, 112.5, 109.4, 109.3, 80.1, 68.2, 65.7, 63.0, 55.4, 55.3, 44.7, 28.2, 15.8. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{FN}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 737.2303; found, 737.2300. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 8:92, flow rate = 0.8 mL/min, λ = 254 nm): t_{minor} = 41.0 min, t_{major} = 44.2 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-Butoxycarbonyl)amino)-6-chloro-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3g**). White solid (67.0 mg, 90% yield). Mp 162–163 °C. $[\alpha]_{\text{D}}^{20}$ +19.0 (c 2.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.45–7.43 (m, 2H), 7.37–7.28 (m, 3H), 7.25–7.21 (m, 4H), 7.00 (d, J = 8.4 Hz, 2H), 6.95 (d, J = 8.8 Hz, 2H), 6.90–6.85 (m, 3H), 6.64 (s, 1H), 5.14–5.07 (m, 2H), 4.97–4.84 (m, 2H), 3.86 (s, 3H), 3.84 (s, 3H), 1.61 (s, 3H), 1.29 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.8, 173.8, 167.9, 161.0, 159.9, 153.8, 145.4, 136.5, 135.3, 135.0, 130.3, 128.8, 127.8, 126.7, 126.1, 125.2, 122.4, 117.3, 115.0, 113.9, 109.5, 80.2, 68.1, 65.0, 63.2, 55.4, 55.3, 44.7, 28.2, 15.9. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{ClN}_2\text{O}_8\text{SNa}$ $[\text{M} + \text{Na}]^+$, 753.2007; found, 753.2009. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 20:80, flow rate = 0.1 mL/min, λ = 254 nm): t_{minor} = 71.8 min, t_{major} = 83.9 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-6-bromo-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3h**). Light yellow solid (76.5 mg, 96% yield). Mp 148–150 °C. $[\alpha]_{\text{D}}^{20}$ –2.5 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.34 (d, J = 7.2 Hz, 2H), 7.27–7.24 (m, 2H), 7.21–7.18 (m, 1H), 7.15–7.11 (m, 4H), 6.94–6.92 (m, 2H), 6.85 (d, J = 8.4 Hz, 3H), 6.79 (d, J = 8.8 Hz, 2H), 6.70 (s, 1H), 5.04–4.96 (m, 2H), 4.87–4.83 (m, 1H), 4.73–4.69 (m, 1H), 3.75 (s, 3H), 3.74 (s, 3H), 1.53 (s, 3H), 1.19 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.7, 173.7, 167.9, 161.1, 159.9, 153.8, 145.5, 136.5, 135.3, 130.3, 128.8, 127.8, 126.7, 125.6, 125.4, 123.0, 117.3, 115.1, 113.9, 112.3, 80.2, 68.2, 65.1, 63.1, 55.4, 55.3, 44.7, 28.2, 15.9. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{N}_2\text{O}_8\text{SBrNa}$ $[\text{M} + \text{Na}]^+$, 797.1502; found, 797.1502. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 5:95, flow rate = 0.5 mL/min, λ = 254 nm): t_{major} = 40.7 min, t_{minor} = 62.4 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-7-chloro-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3i**). Light yellow solid (70.0 mg, 93% yield). Mp 168–169.5 °C. $[\alpha]_{\text{D}}^{20}$ –9.3 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.37 (d, J = 7.2 Hz, 2H), 7.23–7.12 (m, 7H), 7.08–7.05 (m, 2H), 6.89 (d, J = 7.6 Hz, 1H), 6.84 (d, J = 8.8 Hz, 2H), 6.80 (d, J = 8.4 Hz, 2H), 6.74 (t, J = 8 Hz, 1H), 5.34–5.30 (m, 1H), 5.18–5.14 (m, 1H), 5.09–5.03 (m, 2H), 3.75 (s, 3H), 3.73 (s, 3H), 1.47 (s, 3H), 1.20 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.2, 174.6, 167.8, 161.0, 159.9, 153.7, 140.3, 137.6, 136.5, 132.0, 130.7, 130.4, 128.3, 127.3, 127.0, 126.8, 123.3, 122.6, 117.4, 115.2, 115.0, 114.0, 80.2, 68.2, 65.0, 63.0, 55.4, 55.3, 45.7, 28.2, 15.7. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{N}_2\text{O}_8\text{SClNa}$ $[\text{M} + \text{Na}]^+$, 753.2007; found, 753.2003. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 5:95, flow rate = 0.4 mL/min, λ = 254 nm): t_{major} = 47.9 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-7-fluoro-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3j**). Light yellow solid (66.3 mg, 90% yield). Mp 144–146 °C. $[\alpha]_{\text{D}}^{20}$ –3.9 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.50 (d, J = 7.2 Hz, 2H), 7.32–7.29 (m, 2H), 7.26–7.21 (m, 5H), 7.08 (s, 1H), 6.95–6.91 (m, 3H), 6.87–6.79 (m, 4H), 5.08–4.99 (m, 4H), 3.83 (s, 3H), 3.81 (s, 3H), 1.48 (s, 3H), 1.25 (s, 9 H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.4, 173.5, 167.8, 161.0, 159.9, 153.7, 148.4, 146.0, 137.0, 136.5, 130.7, 130.3, 128.4, 128.2, 127.5, 126.7, 123.1, 123.0, 120.0, 117.8, 117.6, 117.4, 115.0, 113.9, 80.1, 68.1, 65.5, 63.0, 55.4, 55.3, 46.1, 28.1, 15.6. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{N}_2\text{O}_8\text{FSNa}$ $[\text{M} + \text{Na}]^+$, 737.2303; found, 737.2303. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 5:95, flow rate = 0.4 mL/min, λ = 254 nm): t_{minor} = 33.4 min, t_{major} = 38.6 min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-7-bromo-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3k**). White solid (72.4 mg, 91% yield). Mp 160–162 °C. $[\alpha]_{\text{D}}^{20}$ +31.0 (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.45 (d, J = 7.2 Hz, 2H), 7.37–7.33 (m, 1H), 7.31–7.26 (m, 5H), 7.24–7.21 (m, 2H), 7.16 (s, 1H), 7.03–7.01 (m, 1H),

6.95–6.89 (m, 4H), 6.78 (t, $J = 8.0$ Hz, 1H), 5.48–5.44 (m, 1H), 5.32–5.28 (m, 1H), 5.17 (m, 2H), 3.85 (s, 3H), 3.84 (s, 3H), 1.59 (s, 3H), 1.30 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.2, 174.8, 167.8, 161.0, 159.9, 153.7, 141.7, 137.5, 136.5, 135.4, 131.0, 130.4, 128.3, 127.1, 126.9, 126.8, 123.7, 123.2, 117.3, 115.0, 114.0, 102.3, 80.2, 68.2, 64.9, 63.0, 55.4, 55.3, 45.4, 28.2, 15.7. HRMS (ESI-TOF): calcd for $\text{C}_{39}\text{H}_{39}\text{BrN}_2\text{O}_8\text{S} [\text{M} + \text{Na}]^+$, 797.1502; found, 797.1499. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 20:80, flow rate = 0.2 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 39.6$ min, $t_{\text{major}} = 58.5$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-2-oxo-7-(trifluoromethyl)indolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3l**). White solid (71.5 mg, 91% yield). Mp 155–157 °C. $[\alpha]_{\text{D}}^{20} +19.0$ (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.57 (d, $J = 8.0$ Hz, 1H), 7.40–7.38 (m, 2H), 7.33–7.27 (m, 5H), 7.23–7.21 (m, 1H), 7.16 (m, 3H), 7.03 (t, $J = 8.0$ Hz, 1H), 6.92 (t, $J = 8.0$ Hz, 4H), 5.26–5.16 (m, 3H), 5.10–5.06 (m, 1H), 3.84 (s, 6H), 1.65 (s, 3H), 1.31 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.1, 175.6, 167.9, 161.1, 159.9, 153.8, 142.6, 136.4, 130.6, 130.4, 128.1, 127.7, 127.6, 127.5, 126.7, 126.6, 126.4, 124.7, 121.9, 117.1, 115.0, 114.0, 112.6, 80.4, 68.3, 63.7, 63.1, 55.4, 55.3, 47.1, 28.1, 15.8. HRMS (ESI-TOF): calcd for $\text{C}_{40}\text{H}_{39}\text{F}_3\text{N}_2\text{O}_8\text{S} [\text{M} + \text{Na}]^+$, 787.2271; found, 787.2274. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 20:80, flow rate = 0.2 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 36.2$ min, $t_{\text{major}} = 50.0$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-3-((Ethoxycarbonyl)amino)-1-methyl-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3m**). White solid (55.5 mg, 91% yield). Mp 142–144 °C. $[\alpha]_{\text{D}}^{20} +7.8$ (c CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.29–7.23 (m, 6H), 7.06 (d, $J = 7.2$ Hz, 1H), 6.95–6.87 (m, 5H), 6.77 (d, $J = 8.0$ Hz, 1H), 5.22–5.14 (m, 2H), 3.88 (q, $J = 6.4$ Hz, 2H), 3.82 (s, 6H), 3.21 (s, 3H), 1.54 (s, 3H), 1.10 (m, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.2, 173.4, 168.0, 161.0, 159.9, 154.6, 145.0, 136.5, 130.4, 129.7, 127.1, 126.8, 124.2, 122.8, 117.5, 115.0, 114.0, 108.1, 68.1, 65.8, 62.7, 61.0, 55.4, 55.3, 26.6, 15.5, 14.3. HRMS (ESI-TOF): calcd for $\text{C}_{31}\text{H}_{33}\text{N}_2\text{O}_8\text{S} [\text{M} + \text{Na}]^+$, 615.1771; found, 615.1771. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 15:85, flow rate = 0.5 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 22.7$ min, $t_{\text{major}} = 39.7$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-3-((Benzylloxy)carbonyl)amino)-1-methyl-2-oxoindolin-3-yl)-3-((4-methoxyphenyl)thio)-2-methyl-3-oxopropanoate (**3n**). White solid (60.8 mg, 90% yield). Mp 150–152 °C. $[\alpha]_{\text{D}}^{20} +12.0$ (c CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.43 (s, 1H), 7.33–7.22 (m, 10H), 7.08 (d, $J = 7.2$ Hz, 1H), 6.98–6.93 (m, 3H), 6.87 (d, $J = 8.8$ Hz, 2H), 6.77 (br, 1H), 5.20 (q, $J = 12.0$ Hz, 2H), 4.92 (q, $J = 12.4$ Hz, 2H), 3.85 (s, 3H), 3.82 (s, 3H), 3.21 (br, 3H), 1.56 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 196.1, 173.3, 168.0, 161.0, 159.9, 154.3, 144.9, 136.5, 130.4, 129.8, 128.3, 128.0, 127.9, 126.7, 124.2, 122.8, 117.5, 115.0, 114.0, 108.1, 68.2, 66.9, 65.9, 62.7, 55.4, 55.3, 26.5, 15.5. HRMS (ESI-TOF): calcd for $\text{C}_{36}\text{H}_{34}\text{N}_2\text{O}_8\text{S} [\text{M} + \text{Na}]^+$, 677.1928; found, 677.1933. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 20:80, flow rate = 0.6 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 36.2$ min, $t_{\text{major}} = 46.5$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-3-((4-chlorophenyl)thio)-2-methyl-3-oxopropanoate (**3o**). Yellow solid (65.0 mg, 90% yield). Mp 157–159 °C. $[\alpha]_{\text{D}}^{20} +42.0$ (c 2.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.46 (d, $J = 7.2$ Hz, 2H), 7.39–7.28 (m, 6H), 7.27–7.17 (m, 4H), 7.12 (d, $J = 7.6$ Hz, 1H), 7.06 (s, 1H), 6.94–6.89 (m, 3H), 6.70 (d, $J = 8.0$ Hz, 1H), 5.18 (m, 2H), 5.03–4.84 (m, 2H), 3.84 (s, 3H), 1.56 (s, 3H), 1.28 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 194.7, 173.8, 168.0, 159.9, 153.7, 144.2, 136.2, 135.8, 130.3, 129.5, 129.4, 128.7, 127.8, 127.6, 127.3, 126.8, 125.5, 124.2, 122.7, 114.0, 109.0, 80.0, 68.2, 65.7, 63.3, 55.3, 44.6, 28.2, 15.7. HRMS (ESI-TOF): calcd for $\text{C}_{38}\text{H}_{37}\text{ClN}_2\text{O}_7\text{S} [\text{M} + \text{Na}]^+$, 723.1902; found, 723.1903. HPLC (Chiralcel OD-H, *i*-propanol/*n*-hexane = 8:92, flow rate = 0.5 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 30.8$ min, $t_{\text{major}} = 27.1$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-6-bromo-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-2-((4-methoxyphenyl)thio)carbonyl)butanoate (**3p**). Light yellow solid (68.9 mg, 85% yield). Mp 165–167 °C. $[\alpha]_{\text{D}}^{20} -12.3$ (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.32 (d, $J = 7.2$ Hz, 2H), 7.26–7.22 (m, 3H), 7.20–7.19 (m, 1H), 7.01–7.00 (m, 6H), 6.85 (d, $J = 8.8$ Hz, 2H), 6.78–6.76

(m, 3H), 6.60 (s, 1H), 4.91–4.88 (m, 1H), 4.76–4.73 (m, 1H), 4.65 (m, 2H), 3.77 (s, 3H), 3.74 (s, 3H), 2.65–2.56 (m, 1H), 2.46–2.37 (m, 1H), 1.18 (s, 9H), 0.94 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 197.2, 172.5, 166.7, 160.1, 158.8, 152.8, 144.3, 135.3, 134.4, 129.8, 129.4, 127.7, 126.8, 126.6, 125.3, 124.5, 124.1, 122.0, 116.6, 114, 0, 112.8, 111.3, 79.3, 67.9, 66.9, 63.4, 54.4, 54.2, 43.7, 27.1, 22.5, 9.3. HRMS (ESI-TOF): calcd for $\text{C}_{40}\text{H}_{41}\text{N}_2\text{O}_8\text{SBrNa} [\text{M} + \text{Na}]^+$, 811.1659; found, 811.1661. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 10:90, flow rate = 0.5 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 35.7$ min, $t_{\text{minor}} = 45.2$ min.

(*R*)-4-Methoxybenzyl 2-((*R*)-1-Benzyl-3-((*tert*-butoxycarbonyl)amino)-2-oxoindolin-3-yl)-2-(((4-methoxyphenyl)thio)carbonyl)pent-4-enoate (**3q**). Light yellow solid (59.6 mg, 80% yield). Mp 152–154 °C. $[\alpha]_{\text{D}}^{20} -64$ (c 1.0 CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.34 (m, 2H), 7.22–7.18 (m, 6H), 7.06–7.00 (m, 4H), 6.91–6.83 (m, 4H), 6.75–6.74 (m, 2H), 6.52–6.51 (m, 1H), 5.65–5.63 (m, 1H), 5.02 (m, 1), 4.96–4.93 (m, 1H), 4.85–4.82 (m, 1H), 4.73–4.70 (m, 2H), 3.75 (d, $J = 4$ Hz, 3H), 3.73 (d, $J = 4.4$ Hz, 3H), 3.29 (m, 1H), 3.12 (m, 1H), 1.16 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3): δ 197.9, 173.4, 167.4, 161.0, 159.7, 153.9, 143.9, 136.3, 136.0, 132.4, 130.8, 130.4, 129.3, 128.6, 128.0, 127.7, 127.5, 126.5, 124.5, 122.4, 119.6, 117.9, 115.0, 113.7, 108.9, 80.1, 68.3, 67.7, 64.6, 55.3, 55.2, 44.6, 35.0, 28.1. HRMS (ESI-TOF): calcd for $\text{C}_{41}\text{H}_{42}\text{N}_2\text{O}_8\text{SNa} [\text{M} + \text{Na}]^+$, 745.2554; found, 745.2558. HPLC (Chiralpak AD-H, *i*-propanol/*n*-hexane = 15:85, flow rate = 0.6 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 35.0$ min, $t_{\text{major}} = 50.1$ min.

■ ASSOCIATED CONTENT

Supporting Information

^1H and ^{13}C NMR spectra of compounds **3a–q** and CIF file of **3h**. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs-joc.5b00302.

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Notes

The authors declare no competing financial interest.

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